

iPSCs? What can we do to help?



Building better human disease models

At Axol Bioscience, we're here to support drug discovery researchers looking to utilize the power of iPSC-derived models.

Like you, we believe that these more human-relevant advanced *in vitro* models have the potential to unlock better, safer therapies for neurodegenerative and cardiovascular diseases.

We've been working with iPSCs in a quality-focused environment for over a decade and have developed a deep understanding of the challenges of this space. Whether it's compound screening on physiologically useful models, iPSC creation, QC and banking, or the large-scale supply of functionally relevant cells, we can help you.

What can you get when outsourcing to Axol?

- High-quality, functionally consistent axoCells™
- A bank of over 70 fully characterized and licensed axoLines™
- Our QC-rich suite of **axoServices**TM, technical iPSC projects
- Cellular assays including multi-electrode array (MEA), 'omics and real-time imaging
- Building iPSC-based **axoModels™** for compound screening

Collaboration is one of our six core company values; we're here to help you by doing the heavy lifting so you can focus on the science with reduced risk, shortened project timelines and improved outcomes.

If you're also a believer in the potential of iPSCs, let's talk.



The 1.

Duncan Borthwick PhD Head of Sales & Marketing

axoCells™







We have developed a catalog of functionally relevant iPSC-derived neurons, neuroinflammatory cells and cardiomyocytes manufactured at our ISO 9001:2015-accredited production facility. Our leading neuronal cell types include: cortical excitatory neurons, striatal neurons, cortical inhibitory interneurons, microglia, astrocytes, sensory neurons and motor neurons.

We also provide high-quality **atrial cardiomyocytes** and **ventricular cardiomyocytes**, as well as made-to-order **myotubes**.

'Ready-to-ship'

| , | • | | | | |
|----------------------------------|-------------------|----------------------------|-----------------|----------------------------|-------------------|
| | axoCells kit code | iPSC-derived cells only | Donor | Disease | Gene/mutation |
| Cortical Excitatory Neurons | ax5111 | ax0111 | Female, 87 | Alzheimer's disease | Apo E4/E4 |
| | ax5112 | ax0112 | Female, 38 | Alzheimer's disease | PSEN1 L286V |
| | ax5113 | ax0113 | Male, 53 | Alzheimer's disease | PSEN1 M146L |
| | ax5114 | ax0114 | Female, 31 | Alzheimer's disease | PSEN1 A246E |
| | ax5115 | ax0015 | Male, newborn | Healthy control | - |
| | ax5116 | ax0016 | Female, newborn | Healthy control | - |
| | ax5118 | ax0018 | Male, 74 | Healthy control | - |
| Striatal neurons | ax3115 | ax0015 | Male, newborn | Healthy control | - |
| | ax3116 | ax0016 | Female, newborn | Healthy control | - |
| | ax3118 | ax0018 | Male, 74 | Healthy control | - |
| | ax3211 | ax0211 | Female, 48 | Huntington's disease | HTT: 50 CAG |
| Cortical Inhibitory Interneurons | - | ax0662 | Male, 40-50 | Healthy control | |
| | - | ax0667 | Male, newborn | Healthy control | |
| Microglia | ax0679 | ax0664 | Male, 40-50 | Healthy control | |
| Astrocytes | - | ax0665 | Male, newborn | Healthy control | |
| Sensory Neurons | | ax0555 | Male, newborn | Healthy control | |
| | ax0157 | ax0055 | Male, newborn | Healthy control | |
| Motor Neurons | | ax0073 | Male, 62 | ALS control (asymptomatic) | C90RF72 >145 G4C2 |
| | | ax0074 | Female, 64 | ALS disease | C90RF72 >145 G4C2 |
| | ax0178 | ax0078 | Male, 74 | Healthy control | |
| Ventricular Cardiomyocytes | ax2500 | ax2508 | Male, 74 | Healthy control | |
| Atrial Cardiomyocytes | - | ax2518 | Male, 74 | Healthy control | |
| Cardiac Fibroblasts | - | ax3039 | Adult | Healthy control | |
| | | | | | |

'Made-to-Order'

| Cell type | Viable cells per vial | ~Time to deliver |
|-------------------------------------|--------------------------|------------------|
| Neurons and neuroinflammatory cells | | |
| Neural Stem Cells (NSCs) | 1.5M | 4 weeks |
| Cortical Excitatory Neurons | 1.5M | 4 weeks |
| Striatal Neurons | 1.5M | 4 weeks |
| Cortical Inhibitory Interneurons | 2M | 4 weeks |
| Microglia | 1M | 17 weeks |
| Astrocytes from iPSCs | 1M | 29 weeks |
| Astrocytes from NSCs | 1M | 16 weeks |
| Sensory Neurons | 0.5M | 10 weeks |
| Motor Neurons | 2M | 13 weeks |
| Cardiac cells | | |
| Ventricular Cardiomyocytes | 2M | 9 weeks |
| Atrial Cardiomyocytes | 1M | 9 weeks |
| Muscle cells | | |
| Myotubes | 1M | 16 weeks |

ISO 9001 quality

Our quality management system is ISO 9001 accredited and demonstrates excellent compliance with ISSCR Standards, demonstrating our commitment to quality and consistency.

What do we mean by 'functional QC'?

Functional QC (fQC) asks simply, are they useful in my assay. By applying fQC we test our iPSC-derived endpoint cells for functional performance with assays that are **biologically relevant** to the cell type and therapy area. This ultimately produces **higher-quality** product that can drive better translational power in robust *in vitro* models.

axoLines™





We've developed a library of over 70 iPSC lines derived from fully-consented patient and healthy control donors. With full licenses and a 50:50 split of male to female donors, our axoLines iPSCs can be used for axoServices custom lab services, made-to-order axoCells projects and specific licensing arrangements.

Key therapy areas include **Alzheimer's Disease**, **Parkinson's Disease**, **ALS**, **Huntington's Disease**, **Friedreich's Ataxia** and **Frontotemporal Dementia**.

5 ALS lines

- We have 5 iPSC lines reprogrammed from patients with ALS (SOD1, TDP43 and C90RF72 mutations):
- CENSOi035-B, an iPSC reprogrammed from a 61-year-old female with ALS (**SOD1** mutation)
- CENSOi018-A, an iPSC reprogrammed from a 62-year-old female with ALS (TDP43 mutation)
- We also have the interesting combination of a disease line derived from 64-year-old female with ALS
 (C90RF72 mutation) and a control line from their sibling, a 62-year-old male with a C90RF72 mutation who
 was asymptomatic at the time of sampling
- We also have a disease control line reprogrammed from a 44-year-old female with a **C90RF72 mutation** (asymptomatic at time of sampling).

7 Alzheimer's Disease lines

- We have 7 iPSC lines reprogrammed from patients with Alzheimer's Disease (APOE4 and PSEN1 mutations) Examples include:
 - An iPSC reprogrammed from an 87-year-old female with Alzheimer's Disease (APOE4 mutation)
 - An iPSC reprogrammed from a 53-year-old male with Alzheimer's Disease (**PSEN1** mutation)
 - For a control line, we recommend the CENSOi004-E line reprogrammed from a 40-50-year-old male.

14 Parkinson's Disease lines

• We have 14 iPSC lines reprogrammed from patients with Parkinson's Disease representing mutations in Ataxin-3, PINK1, PARKIN, PARK2, SNCA and LRRK2

Examples include:

- CENSOi028-A, an iPSC reprogrammed from a 52-year-old female with Parkinson's Disease (PINK1)
- CENSOi030-A, an iPSC reprogrammed from a 54-year-old female with Parkinson's Disease (SNCA)
- For a control line, we recommend CENSOi004-E reprogrammed from a 40-50-year-old male.

6 Huntington's Disease lines

- We have 6 iPSC lines reprogrammed from patients with Huntington's Disease representing mutations in HTT Examples include:
 - CENSOi017-A, an iPSC reprogrammed from a 51-year-old female with Huntington's Disease (HTT)
 - CENSOi053-A, a disease control line reprogrammed from a 64-year-old male who was asymptomatic at the time of sampling (HTT).

We have included the full list overleaf, but you can also visit our axoLines page at https://axolbio.com/axolines/ or scan the QR code. We also have **gene editing and R&D licensing capabilities** for some of our axoLines: contact operations@axolbio.com if you have any queries.

axoLines™

Below you can find a complete list of our iPSC lines:

| Control Lines | Status at time of sampling | Gender | Age of sampling | Source of sampling | iPSC cell line | | |
|--|---|--------|-----------------|--|----------------------------|---|-------------------------------|
| Control | No disease diagnosis | Male | 40-50 | Fibroblasts | CENSOi004-E | | |
| | No disease diagnosis | Male | Newborn | Cordblood | ax7015 | | |
| | No disease diagnosis | Female | Newborn | Cordblood | ax7016 | | |
| | No disease diagnosis | Male | 74 | Pulmonary fibroblast | ax7018 | | |
| Disease Lines ALS | Status at time of sampling | | Age at sampling | Source material | Mutation | Variant Heterozygous D109Y (G>T) mutation | IPSC cell line CENSOi035-B |
| ALS | Patient Asymptomatic (Sibling to | | 61 62 | Fibroblasts Dermal fibroblast | SOD1 C90RF72: >145 G4C2 | Heterozygous D1091 (G>1) mutation | ах7073 |
| | ax7074) | | - | | | | |
| | Patient (Sibling to ax7073) | | 64 | Dermal fibroblast | C90RF72: >145 G4C2 | 100 | ax7074 |
| | Asymptomatic Patient (also with FTD) | | 44 62 | Fibroblasts Fibroblasts | C90RF72 TARDBP | Heterozyogous >100 expanded GGGGCC TARDBP: A382T | CENSOi027-D CENSOi018-A |
| Alzheimer's disease | Patient (also with 11b) | | 31 | Dermal fibroblast | PSEN1 A246E | ApoE: E3/E4 | ax7114 |
| | Patient | | 53 | Dermal fibroblast | PSEN1 M146L | ApoE: E2/E3 | ax7113 |
| | Patient Patient | | | Dermal fibroblast Dermal fibroblast | PSEN1 L286V ApoE | ApoE: E3/E3 ApoE: E4/E4 | ax7112 ax7111 |
| | Patient | | 59 | PBMCs | АроЕ | ApoE: E3/E3 | CENSOi070-A |
| | Patient | Male | 60 | PBMCs | ApoE | ApoE: E3/E4 | CENSOi074-A |
| Behavioural variant frontotemporal | Patient | | 52 | PBMCs | ApoE MAPT | ApoE: E3/E4 Heterozygous c.1920+16C>T in the MAPT gene | CENSOi077-C CENSOi059-A |
| dementia (bvFTD) | Patient | | 65 | Fibroblasts | | ,, | |
| , | Patient Patient | | | Fibroblasts Fibroblasts | MAPT MAPT | Heterozygous c.1920+16C>T Heterozygous c.1920+16C>T | CENSOi060-B CENSOi061-A |
| Charcot-Marie-Tooth Disease, Type | | | 27 | PBMCs | FIG4 | Heterozygous c.122T>C p.(Ile41Thr) exon 2 | CENSOi068-A |
| 4J (CMT4J) | | | | | | | |
| Dentatorubral-pallidoluysian atrophy (DRPLA) | Patient | Male | 16 | Fibroblasts | ATN1 | Heterozygous repeats 1 allele (approximately 13 repeats) in the normal range and one allele (approximately 66 repeats) | CENSOi054-B |
| | Patient | Male | 51 | Fibroblasts | ATN1 | Heterozygous pathogenic repeat expansion in the | CENSOi055-A |
| | Asymptomatic | Female | 45 | Fibroblasts | ATN1 | ATN1 gene (17 ± 1/ 57 ± 2 CAG repeats) ATN1:12±1/13±1 CAG repeats (normal range) | CENSOi058-A |
| Epilepsy | Patient | | 5mo | Dermal fibroblast | Undetermined | Unknown | ax7411 |
| Epilepsy Friedreich's Ataxia | Patient | | 5mo 21 | Fibroblasts | FXN | Homozygous repeat expansion (>75 GAA repeats) | |
| | Patient | | 17 | Fibroblasts | FXN | Homozygous repeat expansion in the FXN gene | CENSOi044-A |
| | Patient | Male | 23 | Fibroblasts | FXN | (>75 GAA repeats) Two pathogenic expanded alleles in the FXN gene | CENSOi048-A |
| | Patient | Male | 34 | Fibroblasts | FXN | (≥66 GAA repeats) Homozygous repeat expansion (>75 GAA repeats) | CENSOi050-A |
| | Asymptomatic | Male | 44 | Fibroblasts | FXN | Homozygous 8±1 GAA repeats | CENSOi039-A |
| | Asymptomatic | Female | Unknown | Fibroblasts | FXN | Heterozygous repeat expansion (>75 GAA repeats) | CENSOi038-A |
| Frontotemporal dementia (FTD) | Patient | | 54 | PBMCs | MAPT | Heterozygous MAPT, c.1920+16C>T | CENSOi069-B |
| | Patient | | 42 | PBMCs | MAPT | Heterozygous MAPT, c.1920+16C>T | CENSOi072-A |
| | Patient Asymptomatic | | 35 46 | PBMCs Fibroblasts | MAPT MAPT | Heterozygous MAPT, c.1920+16C>T Heterozygous c.1920+16C>T | CENSOi076-A CENSOi062-A |
| | Patient | | 31 | Fibroblasts | Progranulin | R493X | CENSOi025-A |
| | Patient | | 65 | Fibroblasts | Progranulin | C31FS | CENSOi032-A |
| FTD, Paget's Disease | Patient Patient | | 43 42 | Fibroblasts Fibroblasts | VCP VCP | Heterozygous VCP R155C (CGT>TGT) Heterozygous VCP R191Q c.572 G>A | CENSOi042-A CENSOi043-A |
| Huntington's Disease | Patient | | 16 | Fibroblasts | нтт | 1 allele within the intermediate range (approximately 28 repeats) and 1 alelle within the expanded range (approximately 66 repeats) | CENSOi052-A |
| | Patient | | | Dermal Fibroblast | НТТ | 50 CAG repeat | ax7211 |
| | Patient | | 40-50 51 | Fibroblasts Fibroblasts | нтт нтт | 18/39 CAG repeats in HTT One HTT allele with CAG repeats in normal range, | CENSOi011-D CENSOi017-A |
| | Patient | remale | 51 | FIDIODIASIS | пп | one HTT allele carrying approximately 42 CAG repeats | CENSUIU I 7-A |
| | Patient | Female | 7 | Fibroblasts | НТТ | One HTT allele with CAG repeats in normal range, one HTT allele carrying approximately 127 CAG repeats | CENSOi019-B |
| | Asymptomatic | Male | 64 | Fibroblasts | нтт | | CENSOi053-A |
| Mucolipidosis IV (ML4) | Patient | | 10 | PBMCs | MCOLN1 | Homozygous MCOLN1, c.406-2A>G | CENSOi064-A |
| | Patient Patient | | 45 7 | PBMCs PBMCs | MCOLN1 MCOLN1 | Heterozygous, MCOLN1, c.406-2A>G Heterozygous, MCOLN1, c.694A>C p.(Thr232Pro); | CENSOI065-B |
| | Patient | wate | , | PBMCS | MCOENT | Heterozygous, MCOLNT, C.694A>C p.(TIII232P10), Heterozygous, MCOLNT, c.785T>C p.(Phe262Ser) | CENSUIU00-A |
| Mass Habala Process | Asymptomatic | | 36 | Fibroblasts | MCOLN1 | Heterozygous MCOLN1, c.785T>C | CENSOi063-A |
| Nasu-Hakola disease | Patient | | 41 | PBMCs | TREM2 | Homozygous TREM2, c.150G>T p.(Trp50Cys) | CENSOi073-A |
| | Patient | | 69 | PBMCs | TREM2 | Heterozygous TREM2, c.150G>T p.(Trp50Cys) | CENSOi075-B |
| | Patient | | 68 | PBMCs | TREM2 | Heterozygous TREM2, c.150G>T p.(Trp50Cys) | CENSOi078-A |
| Parkinsonism/Machado-Joseph | Patient | | 50 | Fibroblasts | ATXN3 | 14/69 CAG repeats in ATXN3 gene | CENSOi021-A |
| disease/ (SCA3) | Patient Patient | | 53 41 | Fibroblasts Fibroblasts | ATXN3 ATXN3 | 26/70 CAG repeats in ATXN3 25/69 CAG repeats | CENSOi022-A CENSOi033-A |
| | Patient | Male | 50 | Fibroblasts | ATXN3 | 29/66 CAG repeats | CENSOi034-A |
| Deukinaania Diaaa | Patient | | 22 | PBMCs | ATXN3 | 14/75 CAG repeats | CENSOi071-A |
| Parkinson's Disease | Patient | | 48 | Fibroblasts | PINK1 & PARKIN | PINK1: Tryp90Leufsx12 PARKIN: Arg275Trp | CENSOi023-A |
| | Patient | Male | 75 | Fibroblasts | PINK1 & PARK2 | Heterozygous PARK2 variant c.823C>Tp.(Arg275Trp). Heterozygous PINK1(W90Lfsx12) (Tryp90Leufsx12) | CENSOi024-A |
| | Patient | Male | 58 | Fibroblasts | PINK1 & PARK2 | Homozygous c.736C>T;p.Arg246 X mutation in the PINK 1 gene Heterozygous deletion encompassing exon 4 to 6 in the PARK2 gene | CENSOi026-C |
| | Patient | | 52 | Fibroblasts | PINK1 | PINK1: Homozygous p.Tryp90Leufsx12 | CENSOi028-A |
| | Patient Patient | | 54 54 | Fibroblasts Fibroblasts | SNCA SNCA | A53T A53T | CENSOi030-A CENSOi031-A |
| | Patient Patient | | 54 48 | Fibroblasts Fibroblasts | SNCA SNCA | Heterozygous G51D (G>A) mutation | CENSOI031-A CENSOI045-A |
| | Patient | Male | 68 | Fibroblasts | LRRK2 | Heterozygous LRRK2(WT/G2019S) | CENSOi046-A |
| SCA2 | Patient | | | Fibroblasts Fibroblasts | LRRK2 ATXN2 | c.4321C>T (p.Arg1441Cys) 22/38 CAG repeats in ATXN2 | CENSOI047-A |
| SCA2 SCA7 | Patient Patient | | 41 61 | Fibroblasts Fibroblasts | ATXN7 | 10/39 CAG repeats in ATXN2 | CENSOi020-A CENSOi036-A |
| Spinal and bulbar muscular atrophy | Patient | | 66 | Fibroblasts | AR | Hemizygous expansion (47±1 CAG repeats) | CENSOi040-A |
| (SBMA) | | | | | | | |
| Spinocerebellar ataxia type 2 (SCA2) | Patient | Male | 58 | Fibroblasts | ATXN2 | 1 CAG repeat allele within the normal range (approximately 22 repeats) and one allele within the SCA2 affected range (approximately 36 | CENSOi051-B |
| Spinocerebellar ataxia type 2 (SCA2) | Patient | Female | Unknown | Fibroblasts | ATXN2 | repeats) One CAG repeat allele within the nromal range (approximately 22 repeats) and one allele within the SCA2 affected range (approximately 39 | CENSOi057-B |
| Spinocerebellar ataxia type 6 (SCA6) | Patient | Female | 63 | Fibroblasts | CACNA1A | repeats) 1 CAG repeat allele within the normal range and one allele within the SCA6 affected range (approximately 23 repeats) | CENSOi056-A |

axoServices™





With over a decade of expertise, Axol Bioscience is established as the **first choice** for professional, QC-rich delivery of outsourced iPSC-related services, working always to our values of performance and transparency.

With a mantra of "do it once, and do it well", we've developed a comprehensive suite of custom lab services. Leave us to do the "heavy lifting", while you get on with the science.

Technical Services

| Service | Deliverable | Timelines |
|--------------------|---|---|
| Reprogramming | iPSCs with QC to demonstrate viability and pluripotency markers, with no adventitious viruses/mycoplasma. | From 21-27 weeks (includes 4 weeks standard QC) |
| Gene-editing | Gene-edited iPSCs with QC. As above plus iPSCs tested for robustness to nucleofection and subcloning, screened for PCR and sequencing to ensure correct targeting, and expanded into banks. | From 28 weeks or longer depending on complexity |
| Differentiation | Vials of frozen iPSC-derived cells with functional QC. Flow cytometry of iPSCs to confirm pluripotency-associated marker expression. On derived cells, ICC to confirm expression lineage-specific markers, checks for contaminants and viability testing. | From 4 weeks depending on cell type |
| Model building | A collaborative iPSC-fueled model that has been built and validated with measurable endpoints and controls. | Custom project |
| Compound screening | Small-scale pilot experiments to optimize assay selection, with progression to larger-scale screening. Full written report and data sharing. | Custom project |

Cellular Assays

We have a range of in-house and outsourced services to support cellular and model QC and compound screening projects.



Our specialties include:

- Multi-electrode array (MEA)
- · Real-time imaging
- Immunocytochemistry
- Confocal and fluorescence microscopy
- Protein and genomic analysis
- Flow cytometry
- · Plate-based assays

axoModels™





We work collaboratively with biopharma organizations to build, assess, and execute advanced in vitro test systems. Guided by our in-house expertise, these models are powered by functional iPSC-derived cells and offer excellent translational potential for research and compound screening. **Each project is unique**, and we work with clients to develop, assess then use these models.

Core model types

Below you can find core in-house models that we can develop in collaboration with groups looking to harness the power of functionally-relevant axoCells.

| Madel Tymes | Calle used in model | Fyamula massurements newfarmed and their utility |
|-------------------------------|--|---|
| Model Types | Cells used in model | Example measurements performed and their utility We would use multi-electrode array (MEA) and real-time calcium flux imaging to identify |
| ALS v1 (MN) | Healthy vs. ALS patient- derived | hyperexcitability and loss of synchronicity. We would also use real-time plate imaging to monitor neurite structure. |
| Amyotrophic lateral sclerosis | motor neurons | |
| ALS v2 (NMJ) | Healthy vs. ALS motor neurons & skeletal muscle | We would use MEA and real-time calcium flux imaging to identify hyperexcitability and loss of synchronicity. We would also use real-time plate imaging to monitor neurite structure and measure muscle contractility via MEA. |
| Amyotrophic lateral sclerosis | | incasare master contracting via MEA. |
| Pain | Sensory neurons | We would look for changes in expression levels of key TRP and NaV channels via RNAseq and immunofluorescence. We would also use MEA and calcium imaging to look for evidence of toxicology, pharmacology, habituation to treatments, measuring parameters that include neuronal activity, capsaicin response, desensitization and tachyphylaxis. We could also use high-content imaging to monitor neurite structure, looking for changes in neurite structure indicating toxicology. |
| Alzheimer's Disease | Spontaneous Alzheimer's Disease (sAD) vs control- derived cortical excitatory neurons, cortical inhibitory neurons, astrocytes & microglia | We would use proteomics to assess cytokine release, using proteomics platforms that measure 30-40 cytokines per well. We would also use genomics to assess RNAseq / genome open reading frame RNA analysis from 96-well array data. High-content imaging would enable us to assess phagocytosis/spontaneous neural activity / high-content imaging in 96-well/384-well format. |
| Neuroinflammation | Healthy vs. AD patient- derived microglia and cortical neurons | We would use high-content imaging to assess phagocytosis, cytokine release, chemotaxis and cortical firing – demonstrating immune response and dysfunction of microglia in neurodegeneration. MEA can be used to measure neuronal activity in co-culture models. |
| Huntington's Disease (HD) | Healthy vs. HD patient- derived striatal neurons | We would use MEA and real-time calcium flux imaging to measure neuronal/calcium channel activity respectively, to identify hyperexcitability and loss of synchronicity in disease model. We would also use high-content imaging to monitor neurite structure (fewer, shorter neurites would indicate neurodegeneration) and PCR/blotting to assess the genomic stability of CAG repeats – assessing disease phenotype with compound treatment effect. |
| Parkinson's Disease (PD) | Healthy vs. PD patient- derived dopaminergic neurons | We would use MEA and real-time calcium flux imaging to measure neuronal/calcium channel activity respectively, to identify hyperexcitability and loss of synchronicity in disease model. We would also use high-content imaging to monitor neurite structure (fewer, shorter neurites would indicate neurodegeneration) |

For more information, scan the QR code on this page or email us at operations@axolbio.com.



Collaborative at heart.

We believe in the power and potential of human iPSCs. If you share this belief, let's work together for mutual success.

We work collaboratively with charitable, academic, regulatory and industrial partners to create ambitious and novel *in vitro* platforms that drive drug discovery forward.

So we ask you:

iPSCs? How can we help?

Learn more.

Access eBooks, posters, papers and protocols at:

www.axolbio.com



Interested? Let's talk.

operations@axolbio.com